

## <sup>199</sup>Hg NMR Correlations in Methylmercury(II) Complexes of Nucleic Acid Constituents and Their Analogs

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The known toxic effects of organomercury compounds are often attributed to the formation of mercury–sulfur bonds with sulfhydryl functions in aminoacids and proteins but may also be due, in part, to their interactions with nucleic acid constituents [1]. <sup>199</sup>Hg NMR (<sup>199</sup>Hg has  $I = \frac{1}{2}$ , a natural abundance of 16.9%, and a sensitivity of 1.4%) has been used as a probe for various interactions of protein constituents with organomercury compounds in solution [2]. <sup>199</sup>Hg NMR has also been used to study the interactions of CH<sub>3</sub>Hg(II) with substituted pyridines, bipyridyls, pyrazoles and related organic compounds [3]. To our knowledge, no report on the use of <sup>199</sup>Hg NMR as a probe for CH<sub>3</sub>Hg(II)–nucleic acid interactions has yet appeared in the literature. As an extension of our <sup>1</sup>H and <sup>13</sup>C NMR studies [4] of the complexes formed by the interactions of CH<sub>3</sub>Hg(II) and nucleic acid constituents, the <sup>199</sup>Hg NMR spectra of these complexes have been determined in (CD<sub>3</sub>)<sub>2</sub>SO. Our results (Fig. 1) indicate that the <sup>199</sup>Hg chemical shifts are characteristic of both the nature of ligand centres (L) bonded to methylmercury(II) (L–HgCH<sub>3</sub>) and the ligand structural type. For ease of discussion one can divide the complexes in Fig. 1 into the categories I–VII as described below. Some of the general relationships which become apparent between <sup>199</sup>Hg chemical shifts and structure are pointed out.

### I. CH<sub>3</sub>Hg(II) Bound to Sulfur of a Pyrimidine Moiety of a Purine Molecule (1a, 1b, 2a, 2b)

The <sup>199</sup>Hg shifts of the protonated cationic complexes 2a and 2b are found upfield compared to the <sup>199</sup>Hg chemical shifts for the corresponding neutral complexes 1a and 1b, and downfield from the values in the protonated cationic complexes 5 and 6 in which CH<sub>3</sub>Hg(II) is bound to an S-centre of imidazole or an imidazole moiety of purine (Group II).

### II. CH<sub>3</sub>Hg(II) Bound to S of Imidazole or an Imidazole Moiety of a Purine (3, 4, 5, 6)

Complexes 3 and 4 have similar chemical shifts, the small difference ( $\Delta\delta = -6.7$  ppm) being presumably due to the presence of the pyrimidine moiety in 4. The <sup>199</sup>Hg chemical shifts of the 1:1 CH<sub>3</sub>Hg(II) cationic complexes 5 and 6 are found upfield from the <sup>199</sup>Hg chemical shifts for the corresponding neutral complexes 3 and 4.

### III. CH<sub>3</sub>Hg(II) Bound to S of Either the Pyrimidine or the Imidazole Ring of Purine with Simultaneous Binding of CH<sub>3</sub>Hg(II) to N-centres of Pyrimidine or Imidazole Moieties of Purine (7a, 7b, 8, 9)

In the 2:1 cationic CH<sub>3</sub>Hg(II) complexes 7a and 7b, where two CH<sub>3</sub>Hg(II) groups are bound simultaneously at S and N, the <sup>199</sup>Hg chemical shifts occur upfield from the values in the corresponding 1:1 cationic CH<sub>3</sub>Hg(II) complexes 2a and 2b, and even further upfield from the corresponding neutral complexes 1a and 1b. The <sup>199</sup>Hg resonance in 8, in which CH<sub>3</sub>Hg(II) groups are bound to N and S of the imidazole moiety of purine, occurs slightly upfield from that in 7b and slightly downfield from 7a in which CH<sub>3</sub>Hg(II) groups are bound to both pyrimidine and imidazole moieties of purine.

### IV. CH<sub>3</sub>Hg(II) Bound to N of Imidazole or the Imidazole Moiety of Purine (10a, 10b, 11a, 11b, 12a, 12b)

In related imidazole-based complexes, 10a/10b, in which H in the first complex is replaced by the NO<sub>2</sub> group in the second, the <sup>199</sup>Hg resonance of the NO<sub>2</sub>-containing species is upfield from that associated with the corresponding H-containing species. This is probably the result of decreased electron density in the ring due to the electron withdrawing effect of the NO<sub>2</sub> group. As previously (e.g. Groups I and III), the <sup>199</sup>Hg chemical shifts in this series of complexes become more negative on going from a neutral CH<sub>3</sub>Hg(II)–N complex to the corresponding cationic CH<sub>3</sub>Hg(II)–N<sup>+</sup> complex.

### V. CH<sub>3</sub>Hg(II) Bound to N of Pyrimidine or the Pyrimidine Moiety of Purine (13a, 13b, 14a, 14b)

Complexes 13a and 13b, which differ only by a methyl group at C<sub>5</sub>, have similar chemical shifts ( $\Delta\delta = 10.9$  ppm). Also, complexes 14a and 14b, differing by an amino group at C<sub>2</sub>, have similar <sup>199</sup>Hg shifts ( $\Delta\delta = 12.6$  ppm).

### VI. CH<sub>3</sub>Hg(II) Groups Bound Simultaneously to N of Either Imidazole, or the Imidazole/Pyrimidine Moieties of a Purine (15a, 15b, 16a, 16b, 17)

In the 2:1 cationic complexes 15a and 15b, in which two CH<sub>3</sub>Hg(II) moieties are bound simultaneously to the pyrimidine and imidazole moieties

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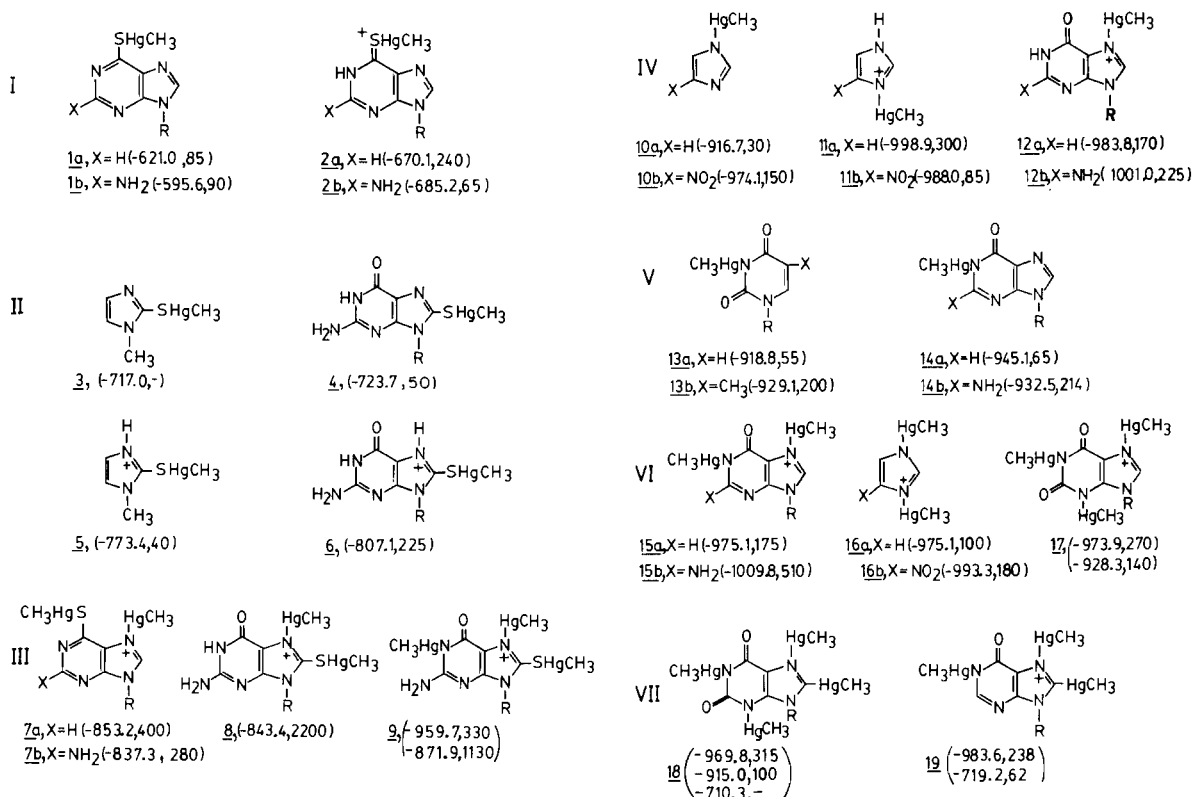


Fig. 1. Structures of the complexes listed under categories I–VII. The data in the parentheses refer to the <sup>199</sup>Hg chemical shifts (negative values, given in ppm relative to (CH<sub>3</sub>)<sub>2</sub>Hg) and the corresponding  $\nu_{1/2}$  values in Hz (see text). Spectra were recorded on a Bruker CXP-200 spectrometer using solutions of the complexes in DMSO-d<sub>6</sub> at a temperature of approximately 335 K.

of purine, the <sup>199</sup>Hg resonances occur upfield by 30.0 and 77.2 ppm, respectively, from the values in the corresponding 1:1 neutral complexes **14a** and **14b**. For complexes **16a** and **16b**, the <sup>199</sup>Hg resonances are also upfield from those of the corresponding neutral complexes **10a** and **10b**. The differences in <sup>199</sup>Hg chemical shift between the pairs **10a/11a**, **10b/11b**, **10a/16a** and **10b/16b**, show that, in this series of complexes, H<sup>+</sup> addition is more effective than CH<sub>3</sub>Hg<sup>+</sup> addition in bringing about the upfield shift.

A noteworthy point with respect to those complexes in groups III–VI, in which there are two CH<sub>3</sub>Hg(II) moieties involved in binding (**7a**, **7b**, **15a**, **15b**, **16a** and **16b**), is that a single averaged <sup>199</sup>Hg resonance is observed as a result of fast exchange, on the NMR time scale, of CH<sub>3</sub>Hg(II) between different sites [5]. This exchange is reflected in a line-width at half-height ( $\nu_{1/2}$ ) in the <sup>199</sup>Hg resonance larger than that observed in the corresponding complex containing only one CH<sub>3</sub>Hg(II) group (e.g. compare  $\nu_{1/2}$  values for pairs **1a/7a**, **1b/7b**, **4/8**, etc., in Fig. 1).

In a number of complexes in which three CH<sub>3</sub>Hg(II) groups are involved in binding to S and N or N sites only, different results are obtained. Thus, in

complex **9**, where two nitrogens and one sulfur are involved in binding to CH<sub>3</sub>Hg(II) groups, two separate <sup>199</sup>Hg resonances are observed; one occurring at -959.7 ppm ( $\nu_{1/2}$  = 230 Hz) and assigned to CH<sub>3</sub>Hg(II) bound to N<sub>1</sub>, and a second resonance at -871.9 ppm ( $\nu_{1/2}$  = 1130 Hz) assigned to the CH<sub>3</sub>Hg(II) groups bound to N<sub>7</sub> and S<sub>8</sub>. The second <sup>199</sup>Hg resonance in **9** has a value comparable to that associated with CH<sub>3</sub>Hg(II) groups bound simultaneously to N<sub>7</sub> and S<sub>8</sub> in **8**. In complex **17**, two resonances are observed; one at -973.9 ppm ( $\nu_{1/2}$  = 270 Hz) due to the two CH<sub>3</sub>Hg(II) groups rapidly exchanging between N<sub>3</sub> and N<sub>7</sub> sites, and the other at -928.3 ppm ( $\nu_{1/2}$  = 140 Hz) assignable to N<sub>1</sub>-bound CH<sub>3</sub>Hg(II).

#### VII. CH<sub>3</sub>Hg(II) Groups Bound Simultaneously to N and C of Imidazole and N of Pyrimidine Moieties of Purine (**18**, **19**)

In complex **18**, three separate <sup>199</sup>Hg resonances, at -969.8, -915.0 and -710.3 ppm, are observed. The resonances at -969.8 ( $\nu_{1/2}$  = 315 Hz) and -915.0 ppm ( $\nu_{1/2}$  = 100 Hz) are comparable to the values observed in **17**. The resonance at -710.3 ppm is therefore assigned to C-bound CH<sub>3</sub>Hg(II). In complex **19**, two <sup>199</sup>Hg resonances have been observed,

one at  $-719.2$  ppm ( $\nu_{1/2} = 62$  Hz) assignable to C–HgCH<sub>3</sub>, and the other at  $-983.6$  ppm ( $\nu_{1/2} = 238$  Hz) assignable to N<sub>1</sub>- and N<sub>7</sub>-bound CH<sub>3</sub>Hg(II). The latter resonance is comparable in frequency and width at half-height to the N<sub>1</sub>, N<sub>7</sub>-bound CH<sub>3</sub>Hg(II) resonance in **15a**.

It is seen that the least negative values of <sup>199</sup>Hg chemical shifts are found for complexes containing S-bonded CH<sub>3</sub>Hg(II), while the most negative values of <sup>199</sup>Hg chemical shifts are found for complexes containing N-bonded CH<sub>3</sub>Hg(II). Thus, in this series of complexes, the <sup>199</sup>Hg chemical shift of CH<sub>3</sub>-Hg(II) bound to a ligand is a useful indicator of the nature of the ligand bound *trans* to the Hg–C bond. Comparison with <sup>13</sup>C chemical shift data [4] shows that donor atoms which give rise to substantial <sup>199</sup>Hg shifts also give rise to substantial <sup>13</sup>C chemical shifts; thus there is a linear relationship between these quantities for the complexes described herein. This relationship presumably holds because a strongly bound ligand weakens the Hg–C bond *trans* to it and thus decreases the 'S' character in the Hg–C bond [6].

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